Device for Sorting of Biological Cells

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ABSTRACT

Cells differ by type not only in shape but also in mechanical, electrical, and magnetic properties. Cells can be sorted using these properties. Based on each principle, devices for cell sorting have been designed. This is a technology applied to cell diagnosis and tissue formation promotion. In this research, the advantages and disadvantages of devices have been compared and the direction of future research was discussed. This technology is expected to be applied to the elucidation of cell properties, disease diagnosis, and regenerative medicine.

Keywords: Cell Sorting, Dielectrophoresis, Microfilter and Microchannel.

1. INTRODUCTION

Cell sorting technology is applied to many devices in the biomedical field. Target cells are sorted from a suspension based on the physical and chemical properties of the cells.

Sorting technology is used in regenerative medicine, analysis, etc. It can be applied to the detection of cancer cells.

In this study, previous research on cell sorting [1] was categorized and the direction of future research was discussed. In cell sorting technology, a target sample is obtained from multiple cells.

This technology is expected to be applied to the elucidation of cell properties, disease diagnosis, and regenerative medicine.

2. METHODS FOR SORTING

Magnetic Field

Using an externally applied magnetic field, cells containing magnetic substances are manipulated [2]. This technology is minimally invasive and does not affect the function of cells *in vivo* [3].

Centrifuge

This technique is often used in experiments in biology and biochemistry. It is based on the principle of separating components with different densities using centrifugal force [4]. Since centrifugal force is used, the rotational speed of the centrifuge is important [5]. Because it is minimally invasive, cells can be sorted without damaging them. Large amounts of samples can be processed simultaneously.

Shear Flow Field

The shear flow field technique is utilized in microchannels. Cells are manipulated using the shear force exerted on them by moving fluids in a flow field at different speeds. This is used for the label free cell separation technique [6].

Laser

Laser produces high-intensity light at a specific wavelength. This property is used to select cells and microparticles [7]. This device requires light source equipment. In the case of identification methods using fluorescent substances, there is a possibility of damaging cells.

Filter

Filters are used as part of microfluidic devices [8, 9]. Sorts particles by passing through or blocking particles of a specific size. If clogging occurs, efficiency decreases.

Dielectrophoresis

Dielectrophoresis manipulates cells using an asymmetric electric field [10]. This will be applied as particulate separation technology [11].

It is also attracting attention as a method for separating and sorting cells. Since cells can be manipulated without contact, the risk of damaging cells is low. Previous research has confirmed that cells are not damaged by electric fields [12].

3. CELL SORTING EXAMPLES

Magnetic Field

Cell separation using a magnetic field allows for non-contact operation [13-17]. Physical damage to cells can be minimized. However, it is necessary to attach magnetic beads or magnetic fluids to the cells. It may affect the state and function of cells.

Microchannel

It is efficient because cells are sorted as the fluid moves. In microchannels, gravity, flow around a small cylinder, inertial flow, and viscoelasticity are used. A method appropriate to the characteristics of the cells is selected. Sorting conditions are difficult to generalize.

By taking advantage of concentration gradients and surface effects, highly sensitive reactions and detection on a fine scale are possible. Fractionation can be performed with a small amount of reagent. Because the device is operated in a small flow path, it is necessary to be handled in a contamination-free environment.

Microfilter

Passage of particles is restricted by size and shape.This is often used in simple methods. Cells may be damaged due to physical contact with the filter surface.

Table 2: Study with microfilter; CTC (Circulating Tumor Cells), CAML (macrophage-like cells).

Shape	Size	Cell	Ref
Hole	$15 \mu m$, $10 \mu m$	CTC	$[25]$
Hole	$d = 7 \mu m$, $t = 10 \mu m$	CTC. CAML	$[26]$
Slit	$w = 10 \text{ }\mu\text{m}$, $t = 25 \mu m$	CTC	$[27]$
Hole	$d = 10 \mu m$,	CTC	$[28]$

Dielectrophoresis

Cell separation using dielectrophoresis allows for non-contact and long-distance operation. It is possible to separate cells while minimizing physical damage to cells. It is a method that can be used for various types of cells.

Table 3: Study with dielectrophoresis; 3T3 (embryonic fibroblasts), HFF (Human Foreskin Fibroblasts), MC (fibroblast), RBC (red blood cell), MDCK (epithelial cells from kidney), HRBC (horse red blood cells), hMSC (Mesenchymal Stem Cell), MDA-MB-231 (human breast adenocarcinoma).

Photolithography technology, sputtering technology [36, 37], electron beam lithography technology [38, 39], and inkjet printing technology [40, 41] are used to manufacture electrodes.

4. DISCUSSION

Each method has advantages and disadvantages. The future direction of cell sorting devices has been discussed. In some cases, sorting cells is difficult. There are cases where the characteristics of each cell cannot be utilized in sorting techniques. There are cases where the differences in characteristics between cells are extremely small. Designing a sorting device that can be applied to a wide variety of cells is difficult, and is a future challenge.

Separation methods using microfilters often involve physical contact between cells and filters. Considering the use of cells after fractionation, it is necessary to select a method that is as least invasive as possible.

It is necessary to bind magnetic beads or magnetic fluids to cells in a magnetic field. There is a need to consider the compatibility and stability of magnetic materials with cells.

In the following, cell separation methods using dielectrophoresis and microchannels are focused on. Non-invasive cell separation is possible using these two methods. In dielectrophoresis, cells are passed along with a fluid and separated by an electric field. It is possible to combine a microchannel on the dielectrophoresis channel. By utilizing multiple characteristics of the cells to be sorted, it is possible to develop an efficient and highly selective sorting device. The challenge is to develop smaller, more portable devices and automated separation systems.

A sorting technique that can be applied to multiple different cells is expected. Applications are expected not only in medicine but also in environmental monitoring. Improvements in stability and efficiency are required. Elucidation of the mechanism of cell sorting will lead to new technologies. It is desirable to develop cell sorting techniques that have less impact on cells.

5. CONCLUSION

In this study, conventional research regarding devices for cell sorting was categorized. Future research developments were considered based on current issues. This technology is expected to be applied to the elucidation of cell properties, disease diagnosis, and regenerative medicine.

REFERENCES

- [1] T. Salafi, K. K. Zeming and Y. Zhang, "Advancements in Microfluidics for Nanoparticle Separation", **Lab Chip**, Vol. 17, No. 1, 2016, pp. 11-33. https://doi.org/10.1039/c6lc01045h
- [2] A. R. A. Fattah, S. Ghosh and I. K. Puri, "High Gradient Magnetic Field Microstructures for Magnetophoretic Cell Separation", **J Chromatogr B Analyt Technol Biomed Life Sci.**, Vol. 1027, No. 1 2016, pp. 194-199. https://doi.org/10.1016/j.jchromb.2016.05.046
- [3] M. Hejazian and N. T. Nguyen, "Negative Magnetophoresis in Diluted Ferrofluid Flow", **Lab Chip**. Vol. 15, No. 14, 2015, pp. 2998-3005.

https://doi.org/10.1039/c5lc00427f

- [4] A. Lee, J. Park, M. Lim, V. Sunkara, S. Y. Kim, G. H. Kim, M. H. Kim and Y. K. Cho, "All-in-One Centrifugal Microfluidic Device for Size-Selective Circulating Tumor Cell Isolation with High Purity", **Anal Chem**., Vol. 86, No. 22, 2014, pp. 11349-11356. https://doi.org/10.1021/ac5035049
- [5] Y. Sun and P. Sethu, "Low-stress Microfluidic Densitygradient Centrifugation for Blood Cell Sorting", **Biomedical Microdevices**, Vol. 20, No. 77, 2018. https://doi.org/10.1007/s10544-018-0323-3
- [6] M. G. Lee, J. H. Shin, C. Y. Bae, S. Choi and J. K. Park, "Label-free Cancer Cell Separation from Human Whole Blood Using Inertial Microfluidics at Low Shear Stress", **Anal Chem**. Vol. 85, No. 13, 2013, pp. 6213-6218. (DOI: https://doi.org/10.1021/ac4006149
- [7] P. H Dannenberg, J. Kang, N. Martino, A. Kashiparekh, S. Forward, J. Wu, A. C Liapis, J. Wang and S. H. Yun, "Laser Particle Activated Cell Sorting in Microfluidics", **Lab Chip**. Vol. 22, No. 12, 2022, pp. 2343-2351. https://doi.org/10.1039/d2lc00235c
- [8] P. Wilding, L.J. Kricka, J. Cheng, G. Hvichia, M. A. Shoffner and P. Fortina, "Integrated Cell Isolation And Polymerase Chain Reaction Analysis Using Silicon Microfilter Chambers", **Anal Biochem**. Vol. 257, No. 2, 1998, pp. 95-100. https://doi.org/10.1006/abio.1997.2530
- [9] S. Kumamoto, K. Nakatake, S. Fukuyama, K. Yasuda, Y. Kitamura, M. Iwatsuki, H. Baba, T. Ihara, Y. Nakanishi and Y. Nakashima, "A Dynamically Deformable Microfilter for

Selective Separation of Specific Substances in Microfluidics", **Biomicrofluidics**. Vol. 14, No. 6, 2020, 064113. https://doi.org/10.1063/5.0025927

- [10] B. Techaumnat, N. Panklang, A. Wisitsoraat and Y. Suzuki, "Study on the Discrete Dielectrophoresis for Particle–cell Separation", **Electrophoresis**. Vol. 41, No. 10-11, 2020, pp. 991-1001. https://doi.org/10.1002/elps.201900473
- [11] S. Hashimoto and R. Ono, "Dielectrophoretic Movement of Cell Passing Between Surface Electrodes in Flow Channel", **ASME Journal of Engineering and Science in Medical Diagnostics and Therapy**, Vol. 7, No. 2, 2024, pp. 1-7.
- [12] C. Huang, S-I Han, H. Zhang and A. Han, "Tutorial on Lateral Dielectrophoretic Manipulations in Microfluidic Systems", **IEEE Trans Biomed Circuits Syst**. Vol. 17, No. 1, 2023, pp. 21-32.

https://doi.org/10.1109/tbcas.2022.3226675

- [13] L. Descamps, M. C. Audry, J. Howard, S. Mekkaoui, C. Albin, D. Barthelemy, L. Payen, J. Garcia, E. Laurenceau, D. L. Roy and A. L. Deman, "Self-Assembled Permanent Micro-Magnets in a Polymer-Based Microfluidic Device for Magnetic Cell Sorting", **Cells**. Vol. 10, No. 7, 2021, 1734. https://doi.org/10.1038/s41598-019-45929-y
- [14] H. G. Kye, B. S. Park, J. M. Lee, M. G. Song, H. G. Song, C. D. Ahrberg and B. G. Chung, "Dual-neodymium Magnet-Based Microfluidic Separation Device", **Sci Rep**. Vol. 9, No. 1, 2019, 9502.

https://doi.org/10.1038/s41598-019-45929-y

[15] J. Yoon, Y. Kang, H. Kim, S. R. Torati, K. Kim, B. Lim and C. G. Kim, "Magnetophoretic Micro-Distributor for Controlled Clustering of Cells", **Adv Sci (Weinh)**. Vol. 9, No. 6, 2022, e2103579.

(DOI: https://doi.org/10.1002/advs.202103579

- [16] A. Shamloo and M. Besanjideh, "Investigation of a Novel Microfluidic Device for Label-Free Ferrohydro-dynamic Cell Separation on A Rotating Disk", **IEEE Trans Biomed Eng**. Vol. 67, No. 2, 2020, pp. 372-378. https://doi.org/10.1109/tbme.2019.2913670
- [17] Y. Liu, R. M. S. Vieira and L. Mao, "Simultaneous and Multimodal Antigen-Binding Profiling and Isolation of Rare Cells via Quantitative Ferrohydro-dynamic Cell Separation", **ACS Nano**. Vol. 17, No. 1, 2023, pp. 94-110. https://doi.org/10.1021/acsnano.2c04542
- [18] A. Ozcelik, S. Gucluer and T. Keskin, "Continuous Flow Separation of Live and Dead Cells Using Gravity Sedimentation", **Micromachines** (Basel). Vol. 14, No. 8, 2023, 1570. https://doi.org/10.3390/mi14081570
- [19] H. Shirinkami, G. Wang, J. Park, J. Ahn, Y. Choi and H. Chun, "Red Blood Cell And White Blood Cell Separation Using A Lateral-dimension Scalable Microchip Based on Hydraulic Jump And Sedimentation", **Sensors and Actuators B: Chemical**. Vol 307, 2020, 127412. https://doi.org/10.1016/j.snb.2019.127412
- [20] M. Xavier, S. H Holm, J. P Beech, D. Spencer, J. O Tegenfeldt, R. O C Oreffo and H. Morgan, "Label-free Enrichment of Primary Human Skeletal Progenitor Cells Using Deterministic Lateral Displacement", **Lab Chip**. Vol. 19, No. 3, 2019, pp. 513-523.

https://doi.org/10.1039/c8lc01154k

- [21] A. Sherbaz, B. M K Konak, P. Pezeshkpour, B. Di Ventura and B. E Rapp, "Deterministic Lateral Displacement Microfluidic Chip for Minicell Purification", **Micromachines (Basel)**. Vol. 13, No. 3, 2022, 365. https://doi.org/10.3390/mi13030365
- [22] C. Macaraniag, J. Zhou, J. Li, W. Putzbach, N. Hay and I. Papautsky, "Microfluidic Isolation of Breast Cancer

Circulating Tumor Cells from Microvolumes of Mouse Blood", **Electrophoresis**. Vol. 44, No. 23, 2023, pp. 1859- 1867. https://doi.org/10.1002/elps.202300108

- [23] A. Raj, K. Ramirez, K. M Young, N. Stone, P. Shankles, M. N. R. Ali, A. M. Compton, W. Lam, A. Alexeev and T. Sulchek, "Label-free Microfluidic Isolation of Functional And Viable Lymphocytes from Peripheral Blood Mononuclear Cells", **Biomicrofluidics**. Vol. 17, No. 5, 2023, 054102. https://doi.org/10.1063/5.0161047
- [24] Y. Zhou, Z. Ma and Y. Ai, "Dynamically Tunable Elastoinertial Particle Focusing And Sorting in Microfluidics", **Lab Chip**. Vol. 20, No. 3, 2020, pp. 568-581. https://doi.org/10.1039/c9lc01071h
- [25] T. Sonoda, N. Yanagitani, K. Suga, T. Yoshizawa, S. Nishikawa, S. Kitazono, A. Horiike, K. Shiba, T. Ishizuka, M. Nishio and S. Matsusaka, "A Novel System to Detect Circulating Tumor Cells Using Two Different Size-selective Microfilters", **Anticancer Res**. Vol. 40, No. 10, 2020, pp. 5577-5582. https://doi.org/10.21873/anticanres.14570
- [26] C. M. Tang, P. Zhu, S. Li, O. V Makarova, P. T Amstutz and D. L Adams, "Blood-based Biopsies-clinical Utility Beyond Circulating Tumor Cells", **Cytometry A**. Vol. 93, No. 12, 2018, pp. 1246-1250.
	- https://doi.org/10.1002/cyto.a.23573
- [27] S. Fukuyama, S. Kumamoto, S. Nagano, S. Hitotsuya, K. Yasuda, Y. Kitamura, M. Iwatsuki, H. Baba, T. Ihara, Y. Nakanishi and Y. Nakashima, "Detection of Cancer Cells in Whole Blood Using A Dynamic Deformable Microfilter And A Nucleic Acid Aptamer", **Talanta**, Vol. 228, 2021, 122239. https://doi.org/10.1016/j.talanta.2021.122239
- [28] Y. Cheng, J. Shen, L. Yuan, Y. Yang, X. Shen, H. Qian, L. Yu, R. Li, X. Lv, T. Yan, Y. Li, L. Wang and B. Liu, "A Novel Device to Capture Circulating Tumor Cells: Quantification And Molecular Analysis in Lung Cancer Patients", **J Biomater Appl**. Vol. 35, No. 1, 2020, pp. 49-58. https://doi.org/10.1177/0885328220914408
- [29] H. K. Chu, Z. Huan, J. K. Mills, J. Yang and D. Sun, "Three-Dimensional Cell Manipulation And Patterning Using Dielectrophoresis Via A Multi-layer Scaffold Structure", **Lab Chip**. Vol.15, No. 3, 2015, pp. 920-930. https://doi.org/10.1039/c4lc01247j
- [30] P. Sharbati, A. K Sadaghiani and A. Koşar, "New Generation Dielectrophoretic-Based Microfluidic Device for Multi-Type Cell Separation", **Biosensors (Basel)**. Vol. 13, No. 4, 2023, 418. https://doi.org/10.3390/bios13040418
- [31] M. Hakoda, Y. Wakizaka and Y. Hirota, "Separation of Viable And Nonviable Animal Cell Using Dielectrophoretic Filter", **Biotechnol Prog**. Vol. 26, No. 4, 2010, pp. 1061- 1067. https://doi.org/10.1002/btpr.394
- [32] M. Hakoda, "Development of Dielectrophoresis Separator with An Insulating Porous Membrane Using DC-Offset AC Electric Fields", **Biotechnol Prog**. Vol. 32, No. 5, 2016, pp. 1292-1300. https://doi.org/10.1002/btpr.2330
- [33] H. Song, J. M Rosano, Y. Wang, C. J Garson, B. Prabhakarpandian, K. Pant, G. J Klarmann, A. Perantoni, L. M Alvarez and E. Lai, "Continuous-flow Sorting of Stem Cells And Differentiation Products Based on Dielectrophoresis", **Lab Chip**. Vol. 15, No. 5, 2015, pp. 1320-1328. https://doi.org/10.1039/c4lc01253d
- [34] K. Kikkeri, B. A Kerr, A. S Bertke, J. S Strobl and M. Agah, "Passivated-electrode Insulator-based Dielectrophoretic Separation of Hetero-geneous Cell Mixtures", **J Sep Sci**. Vol. 43, No. 8, 2020, pp. 1576-1585. https://doi.org/10.1002/jssc.201900553
- [35] F. S. Rizi, S. Talebi, M. K D Manshadi and M. Mohammadi, "Separation of Bacteria Smaller Than 4 µm from Other Blood Components Using Insulator-based Dielectrophoresis: Numerical Simulation Approach", **Biomech Model Mechanobiol**. Vol. 22, No. 3, 2023, pp. 825-836. <https://doi.org/10.1007/s10237-022-01683-1>
- [36] D. P Heineck, B. Sarno, S. Kim and M. Heller, "Electrochemical Attack and Corrosion of Platinum Electrodes in Dielectrophoretic Diagnostic Devices", **Anal Bioanal Chem**. Vol. 412, No. 16, 2020, pp. 3871-3880. https://doi.org/10.1007/s00216-020-02607-7
- [37] M. Kamata, Y. Taguchi and Y. Nagasaka, "Design of An Optofluidic Diffusion Sensor by Transient Grating Using Dielectrophoresis", **Opt Express**. Vol. 26, No. 13, 2018, pp. 16970-16983.

https://doi.org/10.1364/oe.26.016970

[38] Y. F. Lin, S. C. Chiu, S. T. Wang, S. K. Fu, C. H. Chen, W. J. Xie, S. H. Yang, C. S. Hsu, J. F. Chen, X. Zhou, Z. Liu, J. Fang and W. B. Jian, "Dielectrophoretic Placement of Quasizero-, One-, And Two-dimensional Nanomaterials into Nanogap for Electrical", **Electrophoresis**, Vol. 33, No. 16, 2012, pp. 2475-2481.

https://doi.org/10.1002/elps.201200145

[39] A. Bezryadin, C. Dekker and G. Schmid, "Electrostatic Trapping of Single Conducting Nanoparticles between Nanoelectrodes", **Appl. Phys. Lett**. Vol. 71, 1997, pp. 1273- 1275.

https://doi.org/10.1063/1.119871

[40] M. Wu, Y. Gao, Q. Luan, I. Papautsky, X. Chen and J. Xu, "Three-dimensional Lab-on-a-foil Device for Dielectrophoretic Separation of Cancer Cells", **Electrophoresis**. Vol. 44, No. 23, 2023, pp. 1802-1809.

https://doi.org/10.1002/elps.202200287

[41] W. Zhang, N. Li, L. Lin, Q. Huang, K. Uchiyama, J. M. Lin, "Concentrating Single Cells in Picoliter Droplets for Phospholipid Profiling on a Microfluidic System", **Small**. Vol. 16, No. 9, 2020, e1903402. https://doi.org/10.1002/smll.201903402